## Proteasome 26S (human), (purified)

High integrity preparation for use in proteasome research.

Highly purified preparation of '26S' proteasomes useful for carrying out *in vitro* protein degradation studies with suitably ubiquitinylated protein substrates.

Citations: 11

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**Ordering Information** 

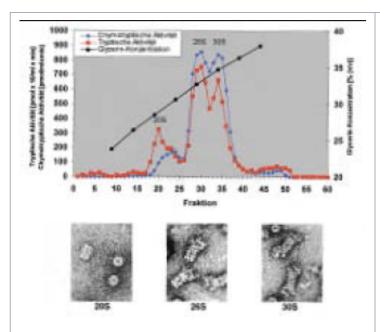
Order Online »

BML-PW9310-0050

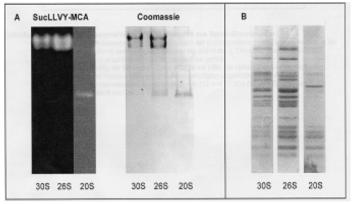
50µg

Manuals, SDS & CofA

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A typical example of a sucrose density gradient separation and purification of 20S, 26S and 30S proteasome derived from human erythrocytes together with electron micrographs of the individual complexes. The protein concentration used for micrography is best at ~2.5µg/mL.



A – Typical results from substrate overlay (carried out in buffer containing ATP and an ATP-regenerating system) and Coomassie staining of non denaturing gel of products isolated according to the purification shown alongside. Protein concentration is ~15-20µg per lane.. B – Coomassie staining of denaturing gel showing presence of proteasomal subunits.

## **Handling & Storage**

**Use/Stability** As indicated on product label or CoA when stored as recommended. When ready for

use the enzyme should be thawed by standing on ice. If the enzymatic acitivity of the 26S proteasome is to be measured, it should be used immediately after thawing since the enzyme complex is labile. After dissociation of the 26S complex the 20S proteasome

activity is relatively stable.

Long Term Storage -80°C

Shipping Dry Ice

## Regulatory Status RUO - Research Use Only

## **Product Details**

Formulation Suspended in TSD buffer (50μg in 10mM TRIS, containing

25mM potassium chloride, 1.1mM magnesium chloride, 0.1mM ethylenediaminetetraacetic acid, 1mM dithiothreitol,

1mM sodium azide, 2mM ATP, pH 7.0, and 35% glycerol).

**Source** Isolated from human erythrocytes. Consists of a high purity

mixture of '26S' proteasomes singly (26S) and doubly (30S) capped with 19S regulatory subunit complexes in the ratio of 40% single cap: 60% double capped at the

time of preparation.

Last modified: May 29, 2024