

# Proteasome 19S Rpt5/S6a subunit polyclonal antibody

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Citations: 15

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## Ordering Information

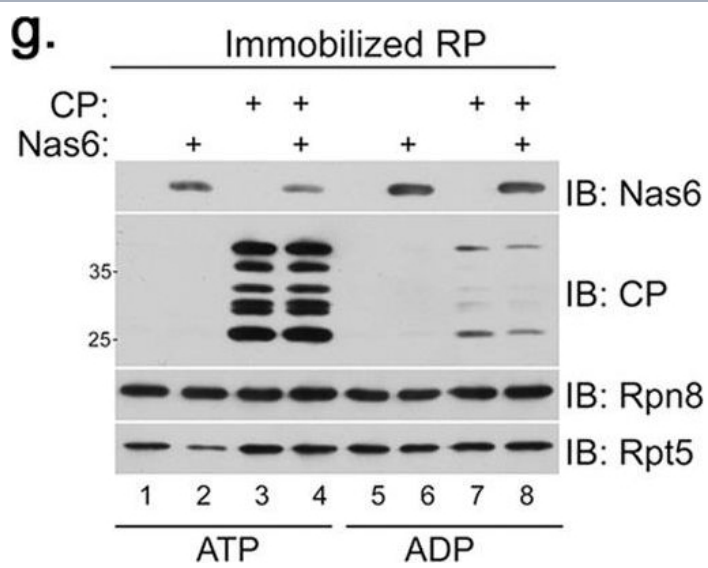
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BML-PW8245-0025	25µl
BML-PW8245-0100	100µl

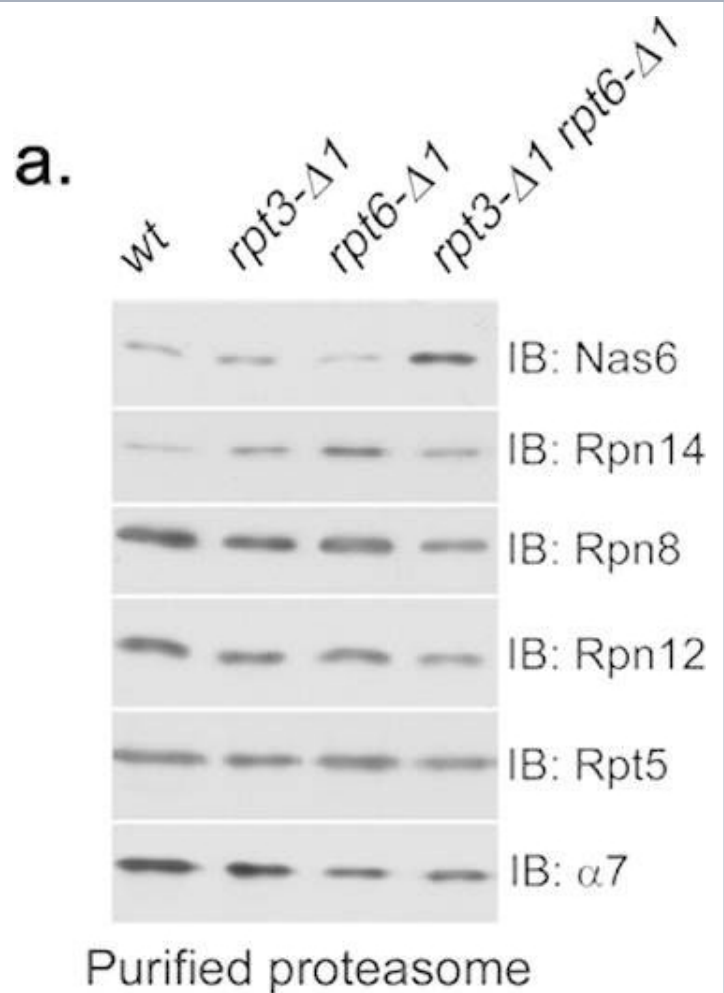
Manuals, SDS & CofA

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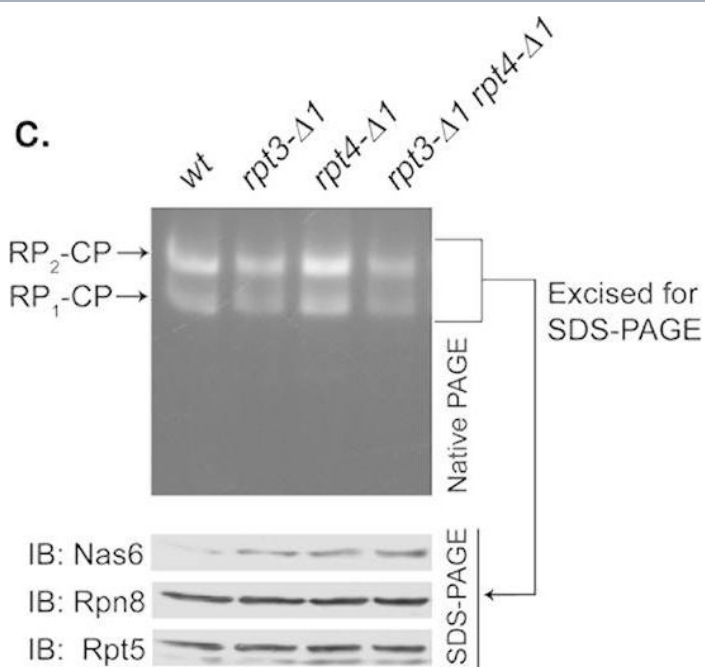




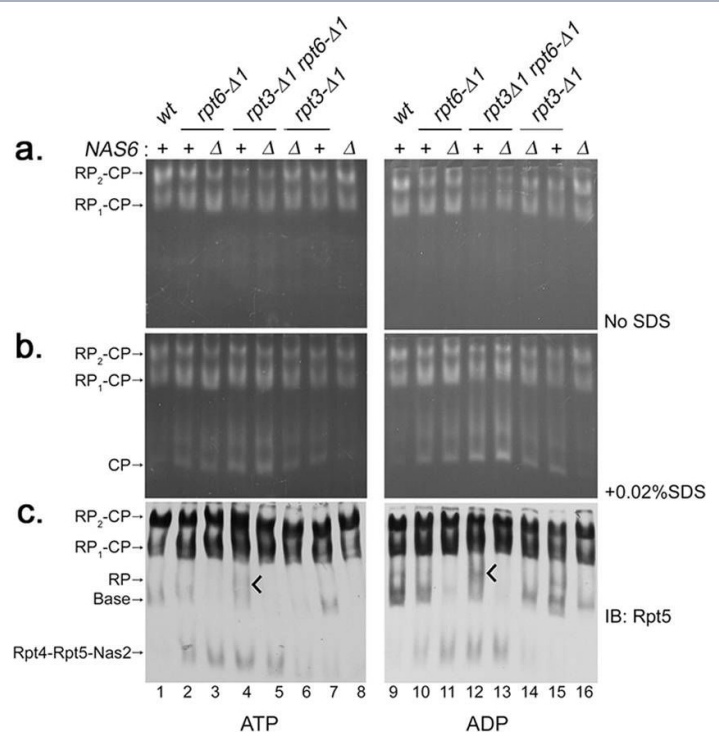
Nas6 maintains the abundance of RP species during Rpt tail-CP interaction. (a,b) Nas6 is not directly responsible for gate opening defects in the proteasome from *rpt3-Δ1rpt6-Δ1* cells. Whole cell lysates (80 μg) were prepared in the presence of ATP (left) or ADP (right) at 2 mM each, and subjected to 3.5% native PAGE. In-gel peptidase assays with LLVY-AMC were performed to assess the gate opening in the proteasome holoenzymes in the absence or presence of 0.02% SDS. (c) Nas6 maintains intracellular levels of RP and base species. The native gels as in (a,b) were subjected to immunoblotting (IB) to detect proteasome holoenzymes and their subassemblies using an antibody to a base subunit Rpt5. RP species become prominent in *rpt3-Δ1rpt6-Δ1* cells under ADP condition (tick mark, lanes 4, 12). (d,e) Proteasome activities (RP2-CP and RP1-CP) of the indicated strains from (a,b) were quantified using ImageJ and normalized to wild-type (% WT) as mean ± SEM (n = 6, ATP; n = 4, ADP). Calculations were performed individually for samples in the absence of SDS, and in the presence of 0.02% SDS. (f) Nas6 promotes RP dissociation from the proteasome in the presence of ADP. Proteasomes were purified with a Protein A tag appended to a lid subunit Rpn1144. Affinity-purified proteasomes (0.6 pmol) were incubated with 1, 5, and 25 fold molar excess of recombinant Nas6 for 30 min at 30 °C in the presence of ATP (2 mM) or ADP (2 mM). The samples were subjected to 3.5% native PAGE and immunoblotting for a base subunit Rpt5 and a lid subunit Rpn8 to detect proteasome holoenzymes and RP species. (g) Nas6 remains bound to the RP when RP-CP interaction is destabilized by ADP. RP was affinity-purified using Rpn11-TEV-ProA from the *nas6Δ* strain and was left on IgG resin. Immobilized *nas6Δ* RP (5 μl) was incubated with purified CP (3 μg) or recombinant Nas6 (0.5 μg), singly and in combination for 30 min at 30 °C, with either ATP or ADP (2 mM). RP-bound material was washed,



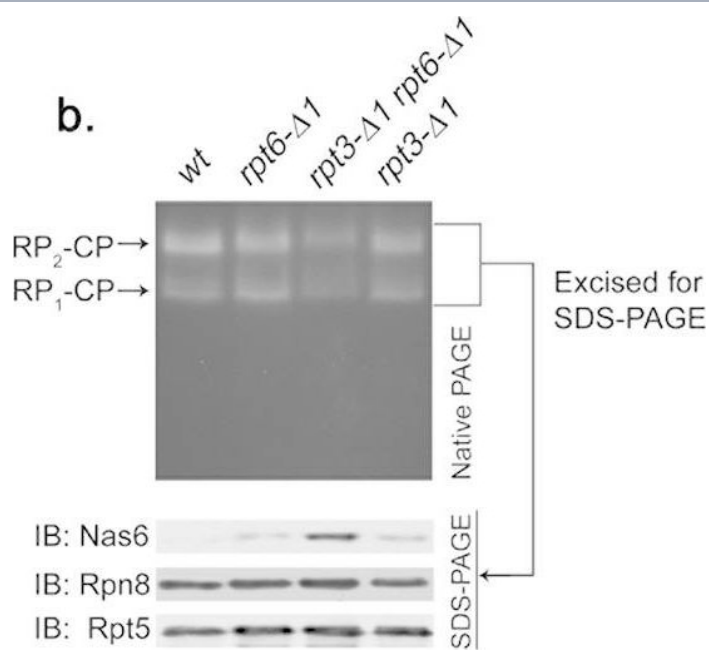
Rpt6 tail binding to the CP is required for Rpt3 tail-mediated release of Nas6 from the proteasome holoenzyme. (a) Increased retention of Nas6 in the proteasomes purified from *rpt3-Δ1rpt6-Δ1* cells. Proteasomes were isolated from indicated strains via a Protein A tag that is appended to CP subunit Pre144. Purified proteasomes (1 μg) were subjected to 12% SDS-PAGE and immunoblotting (IB) for the indicated proteins: Nas6, Rpn14, chaperones; Rpn8, Rpn12, lid subunits; Rpt5, base subunit; α7, CP subunit. Pre1-TEV-ProA tag was chosen to exclude the purification of free RP. Note that Nas6 abundance remains unchanged in these cells (Fig. 1b). (b) Increased retention of Nas6 in the proteasomes in whole cell extracts from *rpt3-Δ1rpt6-Δ1* cells. Whole cell lysates (40 μg) from indicated strains were resolved by 3.5% native gel (top panel). Following proteasome activity assay with LLVY-AMC, the native gel region containing the proteasome holoenzymes (RP2-CP and RP1-CP) was excised into a horizontal strip and directly subjected to 12% SDS-PAGE and immunoblotting for indicated proteins (bottom panel). Rpn8 (a lid subunit) and Rpt5 (a base subunit) serve as loading controls for proteasome levels in the native gel strip. Note that decreased proteasome activities in *rpt3-Δ1rpt6-Δ1* cells are due to gate opening defects (Fig. 2a,b). (c) Nas6 level in the proteasomes from *rpt3-Δ1rpt4-Δ1* cells is comparable to that in the proteasomes from *rpt3-Δ1* cells. Experiments were



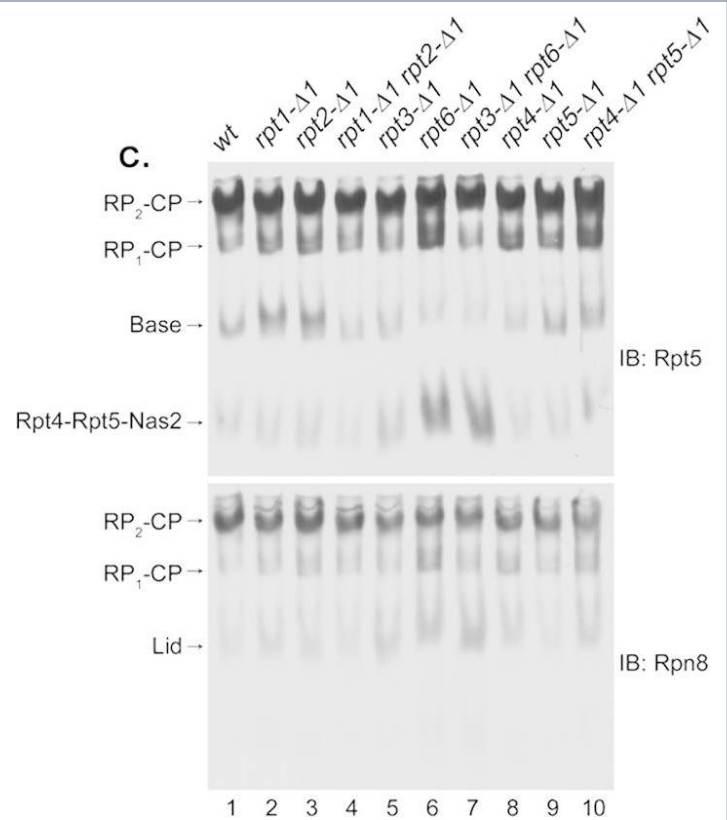
Rpt6 tail binding to the CP is required for Rpt3 tail-mediated release of Nas6 from the proteasome holoenzyme. (a) Increased retention of Nas6 in the proteasomes purified from rpt3- $\Delta$ 1rpt6- $\Delta$ 1 cells. Proteasomes were isolated from indicated strains via a Protein A tag that is appended to CP subunit Pre144. Purified proteasomes (1  $\mu$ g) were subjected to 12% SDS-PAGE and immunoblotting (IB) for the indicated proteins: Nas6, Rpn14, chaperones; Rpn8, Rpn12, lid subunits; Rpt5, base subunit;  $\alpha$ 7, CP subunit. Pre1-TEV-ProA tag was chosen to exclude the purification of free RP. Note that Nas6 abundance remains unchanged in these cells (Fig. 1b). (b) Increased retention of Nas6 in the proteasomes in whole cell extracts from rpt3- $\Delta$ 1rpt6- $\Delta$ 1 cells. Whole cell lysates (40  $\mu$ g) from indicated strains were resolved by 3.5% native gel (top panel). Following proteasome activity assay with LLVY-AMC, the native gel region containing the proteasome holoenzymes (RP2-CP and RP1-CP) was excised into a horizontal strip and directly subjected to 12% SDS-PAGE and immunoblotting for indicated proteins (bottom panel). Rpn8 (a lid subunit) and Rpt5 (a base subunit) serve as loading controls for proteasome levels in the native gel strip. Note that decreased proteasome activities in rpt3- $\Delta$ 1rpt6- $\Delta$ 1 cells are due to gate opening defects (Fig. 2a,b). (c) Nas6 level in the proteasomes from rpt3- $\Delta$ 1rpt4- $\Delta$ 1 cells is comparable to that in the proteasomes from rpt3- $\Delta$ 1 cells. Experiments were carried out as in (b) to assess the level of Nas6 in the proteasome holoenzymes from indicated cells. Rpn8 (a lid subunit) and Rpt5 (a base subunit) serve as loading controls for proteasome levels in the native gel strip. (d) Phenotypic analysis of indicated yeast strains showing that the Rpt3 tail exhibits stronger genetic interaction with the Rpt6 tail than the Rpt4 tail. Four-fold serial dilutions of the indicated cells were spotted onto



Nas6 maintains the abundance of RP species during Rpt tail-CP interaction. (a,b) Nas6 is not directly responsible for gate opening defects in the proteasome from rpt3- $\Delta$ 1rpt6- $\Delta$ 1 cells. Whole cell lysates (80  $\mu$ g) were prepared in the presence of ATP (left) or ADP (right) at 2 mM each, and subjected to 3.5% native PAGE. In-gel peptidase assays with LLVY-AMC were performed to assess the gate opening in the proteasome holoenzymes in the absence or presence of 0.02% SDS. (c) Nas6 maintains intracellular levels of RP and base species. The native gels as in (a,b) were subjected to immunoblotting (IB) to detect proteasome holoenzymes and their subassemblies using an antibody to a base subunit Rpt5. RP species become prominent in rpt3- $\Delta$ 1rpt6- $\Delta$ 1 cells under ADP condition (tick mark, lanes 4, 12). (d,e) Proteasome activities (RP2-CP and RP1-CP) of the indicated strains from (a,b) were quantified using ImageJ and normalized to wild-type (% WT) as mean  $\pm$  SEM (n = 6, ATP; n = 4, ADP). Calculations were performed individually for samples in the absence of SDS, and in the presence of 0.02% SDS. (f) Nas6 promotes RP dissociation from the proteasome in the presence of ADP. Proteasomes were purified with a Protein A tag appended to a lid subunit Rpn1144. Affinity-purified proteasomes (0.6 pmol) were incubated with 1, 5, and 25 fold molar excess of recombinant Nas6 for 30 min at 30  $^{\circ}$ C in the presence of ATP (2 mM) or ADP (2 mM). The samples were subjected to 3.5% native PAGE and immunoblotting for a base subunit Rpt5 and a lid subunit Rpn8 to detect proteasome holoenzymes and RP species. (g) Nas6 remains bound to the RP when RP-CP interaction is destabilized by ADP. RP was affinity-purified using Rpn11-TEV-ProA from the nas6 $\Delta$  strain and was left on



Rpt6 tail binding to the CP is required for Rpt3 tail-mediated release of Nas6 from the proteasome holoenzyme. (a) Increased retention of Nas6 in the proteasomes purified from *rpt3- $\Delta$ 1rpt6- $\Delta$ 1* cells. Proteasomes were isolated from indicated strains via a Protein A tag that is appended to CP subunit Pre144. Purified proteasomes (1  $\mu$ g) were subjected to 12% SDS-PAGE and immunoblotting (IB) for the indicated proteins: Nas6, Rpn14, chaperones; Rpn8, Rpn12, lid subunits; Rpt5, base subunit;  $\alpha$ 7, CP subunit. Pre1-TEV-ProA tag was chosen to exclude the purification of free RP. Note that Nas6 abundance remains unchanged in these cells (Fig. 1b). (b) Increased retention of Nas6 in the proteasomes in whole cell extracts from *rpt3- $\Delta$ 1rpt6- $\Delta$ 1* cells. Whole cell lysates (40  $\mu$ g) from indicated strains were resolved by 3.5% native gel (top panel). Following proteasome activity assay with LLVY-AMC, the native gel region containing the proteasome holoenzymes (RP<sub>2</sub>-CP and RP<sub>1</sub>-CP) was excised into a horizontal strip and directly subjected to 12% SDS-PAGE and immunoblotting for indicated proteins (bottom panel). Rpn8 (a lid subunit) and Rpt5 (a base subunit) serve as loading controls for proteasome levels in the native gel strip. Note that decreased proteasome activities in *rpt3- $\Delta$ 1rpt6- $\Delta$ 1* cells are due to gate opening defects (Fig. 2a,b). (c) Nas6 level in the proteasomes from *rpt3- $\Delta$ 1rpt4- $\Delta$ 1* cells is comparable to that in the proteasomes from *rpt3- $\Delta$ 1* cells. Experiments were carried out as in (b) to assess the level of Nas6 in the proteasome holoenzymes from indicated cells. Rpn8 (a lid subunit) and Rpt5 (a base subunit) serve as loading controls for proteasome levels in the native gel strip. (d) Phenotypic analysis of indicated yeast strains showing that the Rpt3 tail exhibits stronger genetic interaction with the Rpt6 tail than the Rpt4 tail. Four-fold serial dilutions of the indicated cells were spotted onto



Rpt6 tail binding to the CP is crucial for Rpt3 tail-mediated gate opening in the proteasome holoenzyme. (a) Decreased proteasome activity in *rpt3- $\Delta$ 1rpt6- $\Delta$ 1* cells (top panel, RP<sub>2</sub>-CP and RP<sub>1</sub>-CP) is restored upon artificial opening of the CP gate (bottom panel, RP<sub>2</sub>-CP and RP<sub>1</sub>-CP). Whole cell lysates (80  $\mu$ g) from indicated strains were resolved by 3.5% native PAGE. The native gel was incubated with fluorogenic peptide substrate LLVY-AMC to visualize the proteasomes (top panel). Following the imaging of the native gel, the same gel was further incubated with LLVY-AMC in the presence of 0.02% SDS (bottom panel). The addition of SDS is known to open the CP gate<sup>8</sup>. RP<sub>2</sub>-CP and RP<sub>1</sub>-CP are doubly-capped and singly-capped proteasome holoenzymes, respectively. The CP gate within wild-type proteasome holoenzymes is in an open configuration and is not further enhanced by 0.02% SDS<sup>8</sup>. (b) Quantification of relative proteasome activities (RP<sub>2</sub>-CP and RP<sub>1</sub>-CP) from seven independent experiments as in (a) plotted as mean  $\pm$  standard error of the mean (SEM). Values of proteasome activities of the indicated strains were quantified using ImageJ software, and normalized to wild-type to obtain the relative proteasome activities. Calculations were performed individually for samples in the absence of SDS as in top panel from (a), and in the presence of 0.02% SDS as in bottom panel from (a) for each experiment. (c) Impaired RP assembly and defective proteasome holoenzymes in *rpt3- $\Delta$ 1rpt6- $\Delta$ 1* cells. Following proteasome activity assays with the LLVY-AMC as in (a), the native gels were subjected to immunoblotting (IB) to detect the proteasome



## Handling & Storage

Long Term Storage -20°C

Shipping Blue Ice

**Regulatory Status** RUO - Research Use Only

## Product Details

Alternative Name	26S protease regulatory subunit 6A, Tat-binding protein homolog 1, TBP-1
Application	WB
Formulation	Liquid. In PBS containing 10mM sodium azide.
GenBank ID	464859
Host	Rabbit
Immunogen	Recombinant protein corresponding to the N-terminal 100 amino acids of Yta1.
Species Reactivity	Yeast
Specificity	Recognizes the Rpt5/S6a subunit of the 19S regulator complex.
UniProt ID	P33297
Worry-free Guarantee	This antibody is covered by our <a href="#">Worry-Free Guarantee</a>

Last modified: May 29, 2024



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