

Ceramide monoclonal antibody (MID 15B4)

This antibody is covered by our [Worry-Free Guarantee](#).

Citations: 103

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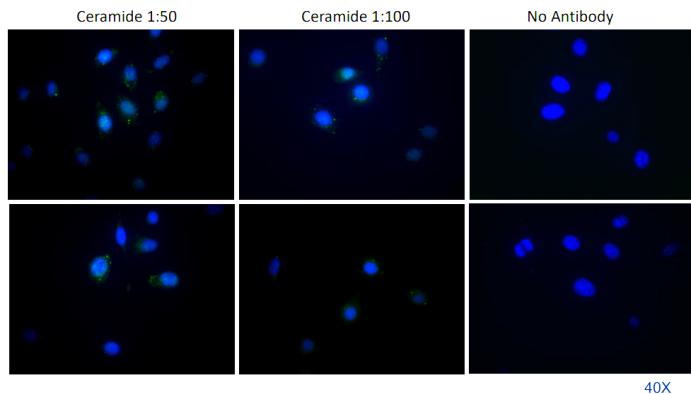
Ordering Information

[Order Online »](#)

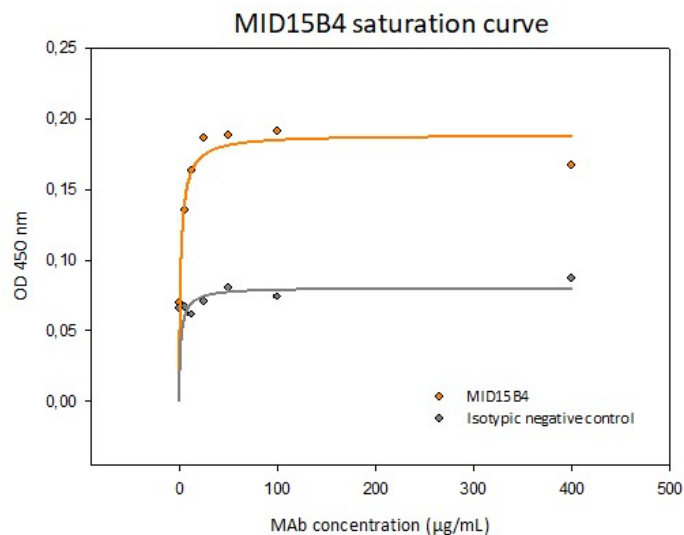
| | |
|------------------|----------|
| ALX-804-196-T050 | 50 tests |
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Manuals, SDS & CofA

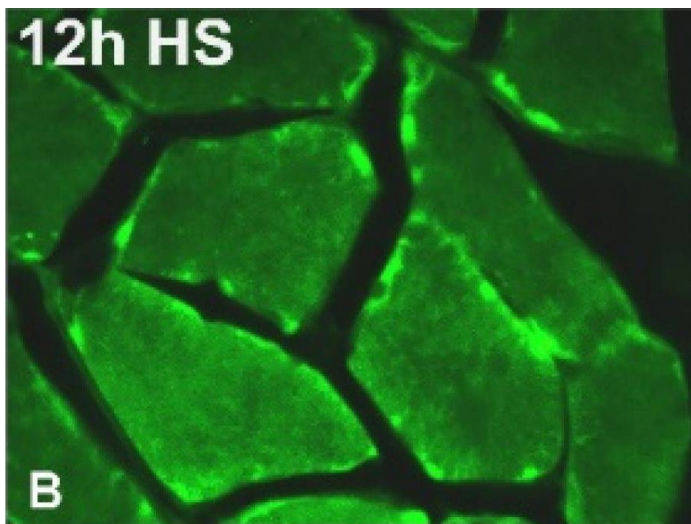
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HeLa cells stained with anti-Ceramide antibody (ALX-804-196-T050) using anti-mouse Alexa Fluor® 488 as secondary ab. Baseline ceramide staining is visible in lipid droplets and stains around cell membranes and membranous organelles.



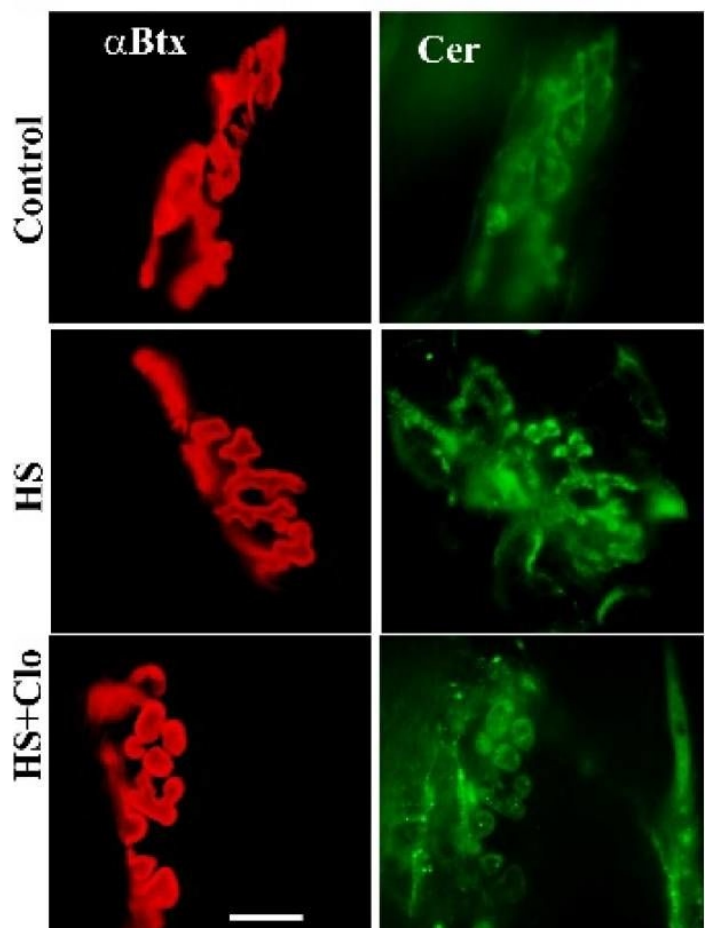
Saturation curve generated for C16-ceramide and negative control by ELISA using Ceramide mAb (MID 15B4)



Expression of immunoreactive ceramide (Cer) on transverse sections of soleus muscle fibers in control rats (A), rats after 12 h of hindlimb suspension (HS) (B) and 12 h of HS with clomipramine (Clo) pretreatment (C). Scale bar—100 μ m. Image (D) represents the negative control. The graph is the quantification of ceramide fluorescence (mean \pm SEM). Data are shown as a percentage of the baseline value (100%) obtained in the control group. ** $p < 0.01$ and *** $p < 0.001$ denote statistically significant differences in comparison with the control value; ## $p < 0.01$ —the difference between non-pretreated and clomipramine-pretreated groups. Control muscles—Control; muscles from HS for 12 h rats—HS; muscles from HS and clomipramine-pretreated animals—HS + Clo. $n = 5$ –6 animals for each group.

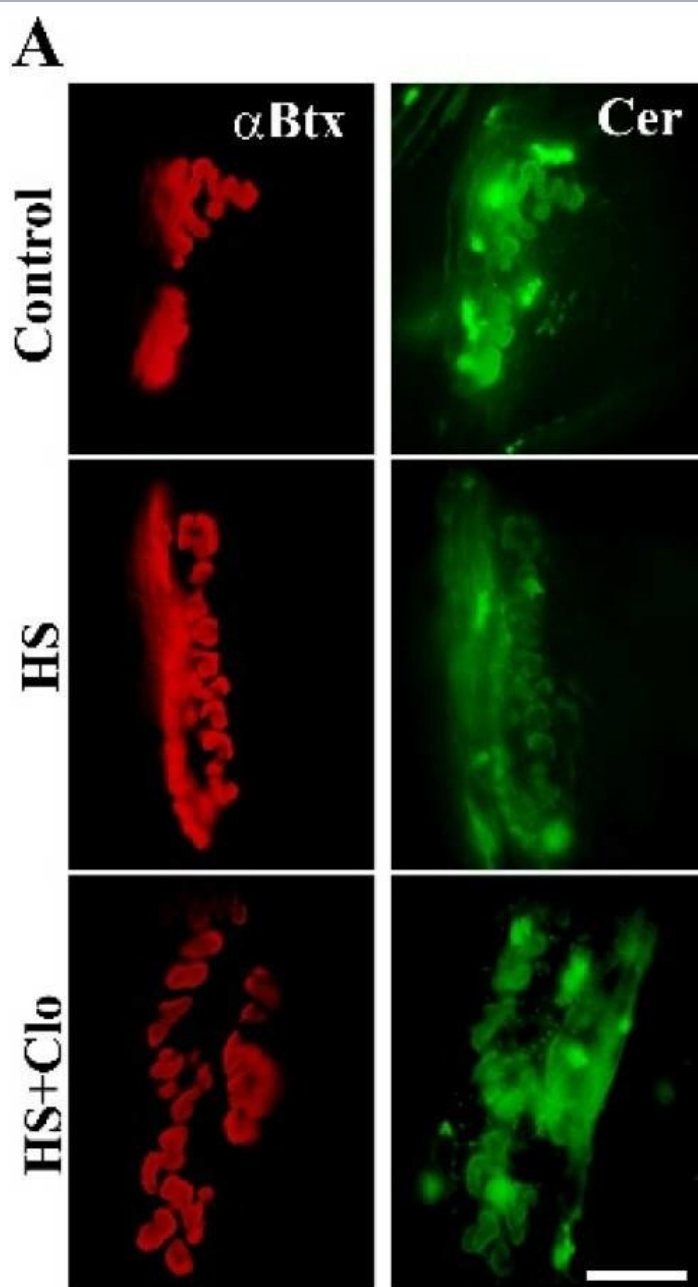
Image collected and cropped by CiteAb under a CC-BY license from the following publication: Changes in Membrane Ceramide Pools in Rat Soleus Muscle in Response to Short-Term Disuse. *Int J Mol Sci* (2019)

A

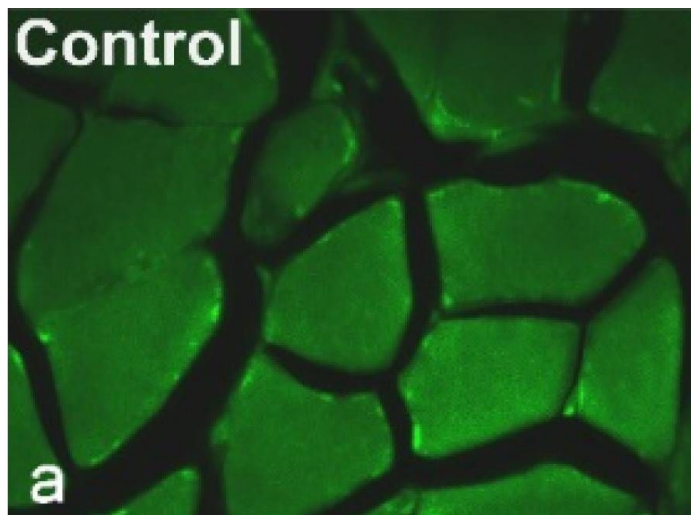


HS increased the immunofluorescent labeling of plasma membrane ceramide: junctional-specific effect of clomipramine therapy. (A) and (B) The fluorescent images of junctional (A) and extrajunctional (B) regions. Membrane Cer was labeled with anti-Cer antibody (green channel) in control, suspended muscle (HS), or suspended muscle of clomipramine-treated rats (HS + Clo). α -Btx (red channel) was used for localization of nicotinic acetylcholine receptors (nAChRs) in postsynaptic membranes. Scale bars—10 μ m. (C) The box plots show the alteration of ceramide immunofluorescent staining (in a.u.) in junctional/extrajunctional compartments in the control, suspended nontreated, and clomipramine-treated muscles. Gray spots represent individual measurements (14–20 measurements per animal and $n = 6$ different animals per group). ** $p < 0.01$, *** $p < 0.001$ are statistically significant differences compared with the corresponding control value. #### $p < 0.001$ is between nontreated and clomipramine-treated groups. Other details are as in Figure 4.

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HS decreased the membrane staining with fluorescent BODIPY FL C5-Ceramide (BODIPY-Cer): influence of clomipramine pretreatment. (A) Junctional regions double-labeled with α -bungarotoxin (α -Btx) and BODIPY-Cer in control, suspended muscle (HS), or suspended muscle of clomipramine-pretreated rats (HS + Clo). Additionally, the green fluorescent spots are visualized in the region surrounding the synaptic zone (perisynaptic region). (B) BODIPY-Cer fluorescence in the extrajunctional regions of the muscle fibers. (A) and (B) scale bars—10 μ m. (C) The box plots indicate the changes in the fluorescent BODIPY-Cer signal in the junctional, extrajunctional, and perisynaptic regions in control, suspended nontreated, and clomipramine-treated muscles. Gray spots represent individual measurements (12–53 measurements per animal and $n = 6$ different animals per group). The measurements were pooled together to obtain the mean values (the central horizontal lines of the boxes). Standard errors (box ranges) and standard deviations (whiskers) are shown. Y-axis—intensity of green fluorescence in a.u. *



Expression of immunoreactive ceramide (Cer) on transverse sections of soleus muscle fibers in control rats (A), rats after 12 h of hindlimb suspension (HS) (B) and 12 h of HS with clomipramine (Clo) pretreatment (C). Scale bar—100 μ m. Image (D) represents the negative control. The graph is the quantification of ceramide fluorescence (mean \pm SEM). Data are shown as a percentage of the baseline value (100%) obtained in the control group. ** $p < 0.01$ and *** $p < 0.001$ denote statistically significant differences in comparison with the control value; ## $p < 0.01$ —the difference between non-pretreated and clomipramine-pretreated groups. Control muscles—Control; muscles from HS for 12 h rats—HS; muscles from HS and clomipramine-pretreated animals—HS + Clo. $n = 5$ –6 animals for each group.

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Handling & Storage

Long Term Storage +4°C

Shipping Blue Ice

Regulatory Status RUO - Research Use Only

Product Details

| | |
|-------------------------------------|--|
| Application | ELISA, Flow Cytometry, ICC, IHC (PS) |
| Clone | MID 15B4 |
| Formulation | Liquid. In PBS, pH 7.2, containing 0.5M sodium chloride, 0.1% BSA and 0.09% sodium azide. |
| Host | Mouse |
| Immunogen | Ceramide (sphingosine-[trans-D-erythro-2-amino-4-octadecene-1,3-diol]) conjugated to BSA. |
| Isotype | IgM |
| Recommendation Dilutions/Conditions | ELISA (1:10)Immunohistochemistry (1:10)Suggested dilutions/conditions may not be available for all applications.Optimal conditions must be determined individually for each application. |
| Source | Purified from ascites by gel filtration on sephacryl S-300. |
| Species Reactivity | Species independent |
| Specificity | Recognizes C16- and C24-ceramide, dihydroceramide, sphingomyelin and phosphatidylcholine in highly artificial lipid overlay test systems. Under more physiological <i>in vitro</i> and <i>in vivo</i> conditions highly specific for ceramide and does not cross-react with sphingomyelin, cholesterol or other phospholipids. |

Technical Info / Product Notes

Cited samples:

[For an overview on cited samples please click here.](#)

Worry-free Guarantee

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Last modified: May 29, 2024



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